

# The Fanconi Anemia Cancer Cell Line Resource

**UM-SCC-01** (*FANCA*<sup>+/+</sup>)

**UM-SCC-01ko** (*FANCA*<sup>-/-</sup>)

**UM-SCC-01ko+Tg** (*FANCA*<sup>+</sup> transgene-complemented from clonally derived *FANCA*<sup>-/-</sup> subline)

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## Description

**Organism:** *Homo sapiens*

**Tissue:** Floor of the mouth

**Disease:** Head and neck squamous cell carcinoma

**Age:** 73Y

**Gender:** male

**HPV status:** negative

**Synonyms:** UM-SCC-1; UM-SCC1; UMSCC1; SCC-1; SCC1

“UM-SCC-01” is a *FANCA*<sup>+/+</sup> sporadic head and neck squamous cell carcinoma (HNSCC) cell line distributed by Millipore Sigma (SCC070). The cell line was isolated from a surgical resection of a recurrent tumor of the floor of the mouth in a 73-year-old male and established at the University of Michigan (1).

“UM-SCC-01ko” is a clonally derived *FANCA*<sup>-/-</sup> cell line generated from UM-SCC-01. The *FANCA* knockout was generated by dual-gRNA cas9-mediated deletion targeting the N-terminus of the *FANCA* gene resulting in a loss of protein expression and function.

“UM-SCC-01ko+Tg” is a clonally derived, *FANCA* transgene-containing (Tg<sup>+</sup>) subline of UM-SCC-01ko in which the *FANCA* transgene has been inserted into the chromosome 4 ‘safe harbor site’ 231 (SHS231) (2).

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## Growth and selection media

UM-SCC-01 was grown in Dulbecco’s Modified Eagle Medium – High glucose (Sigma D5796), 10% fetal bovine serum (FBS, Hyclone Laboratories SH30071.03), 1% penicillin-streptomycin (Gibco 15140122), and 1% non-essential amino acids (Gibco 11140050).

Antibiotic selection of UM-SCC-01ko+Tg cells after thawing and prior to experiments is

recommended. Selection media: 100 ug/ml hygromycin B in complete media.

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## Cryopreservation and thaw protocol

Cells were cryopreserved in freezing medium containing 90% complete media and 10% DMSO.

We recommend thawing cells upon arrival to assess viability, and to do test thaws for viability when freezing liquid nitrogen stocks. Thaw the vial in a 37°C bath for 2-3 minutes before gently transferring to a centrifuge tube. Spin at 1000 RPM before resuspending in pre-warmed complete media. Plate the cells in the culture vessel of choice.

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## Select genomic data

Genomic characterization of the top 12 frequently mutated genes in HNSCC (3).

Gene	TCGA mutfreq (n=276)	Mutation Status
<i>TP53</i>	72%	
<i>FAT1</i>	23%	
<i>CDKN2A</i>	22%	
<i>PIK3CA</i>	21%	
<i>NOTCH1</i>	19%	
<i>KMT2D</i>	18%	
<i>NSD1</i>	10%	
<i>CASP8</i>	9%	Nonsense
<i>AJUBA</i>	6%	
<i>FBXW7</i>	5%	
<i>HRAS</i>	4%	
<i>PTEN</i>	2%	

**Additional genomic characterization can be found in the resource manuscript (4) and following publications (5,6).**

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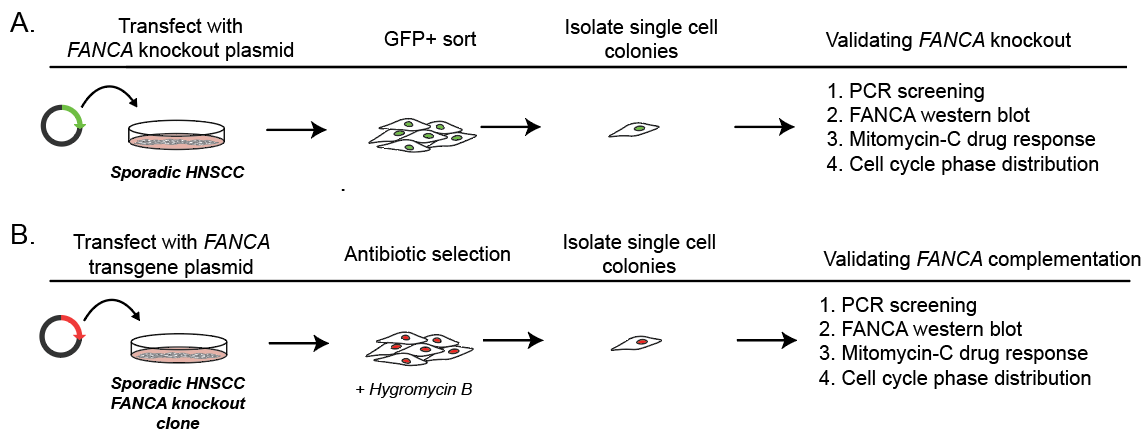
## Quality Control Testing

- The terminal expansion samples of the complete trio set were sent to IDEXX BioAnalytics (Columbia, MO, USA) for STR authentication using their Cell Check 9 Plus service. Cells were simultaneously tested for *Mycoplasma* and interspecies contamination from mouse, rat, African green monkey, and Chinese hamster. Distributed cell lines all had unambiguous STR profile data and were negative for all screened contaminants.
- A Human 9 species-specific STR marker profile was established for the cell line pair. The unique marker results from our recent analysis are listed in STR profiling panels below.
- Test thaws indicate  $\geq 50\%$  survival of cells.

STR: ATCC	STR: IDEXX	STR: IDEXX	STR: IDEXX
<b>UM-SCC-01</b>	<b>UM-SCC-01</b>	<b>UM-SCC-01ko</b>	<b>UM-SCC-01ko+Tg</b>
AMEL: X, Y	AMEL: X	AMEL: X	AMEL: X
CSF1PO: 10,12	CSF1PO: 10,12	CSF1PO: 10,12	CSF1PO: 10,12
D13S317: 8,11	D13S317: 8,11	D13S317: 8,11	D13S317: 8,11
D16S539: 12,13	D16S539: 12,13	D16S539: 12,13	D16S539: 12,13
D5S818: 10,13	D5S818: 10,13	D5S818: 10,13	D5S818: 10,13
D7S820: 11,12	D7S820: 11,12	D7S820: 11,12	D7S820: 11,12
TH01: 9,12	TH01: 9,12	TH01: 9,12	TH01: 9,12
TPOX: 8,11	TPOX: 8,11	TPOX: 8,11	TPOX: 8,11
vWA: 15,18	vWA: 15,18	vWA: 15,18	vWA: 15,18

Note: Previous reports have identified differences in the Amelogenin marker for UM-SCC-01 that are dependent on the passage number (7). The UM-SCC-01 parent cell line included in the FA-CCLR matches the reported STR profile for the long-term passage line. **More extensive STR profiling data can be found the following publications (8,9).**

## General workflow

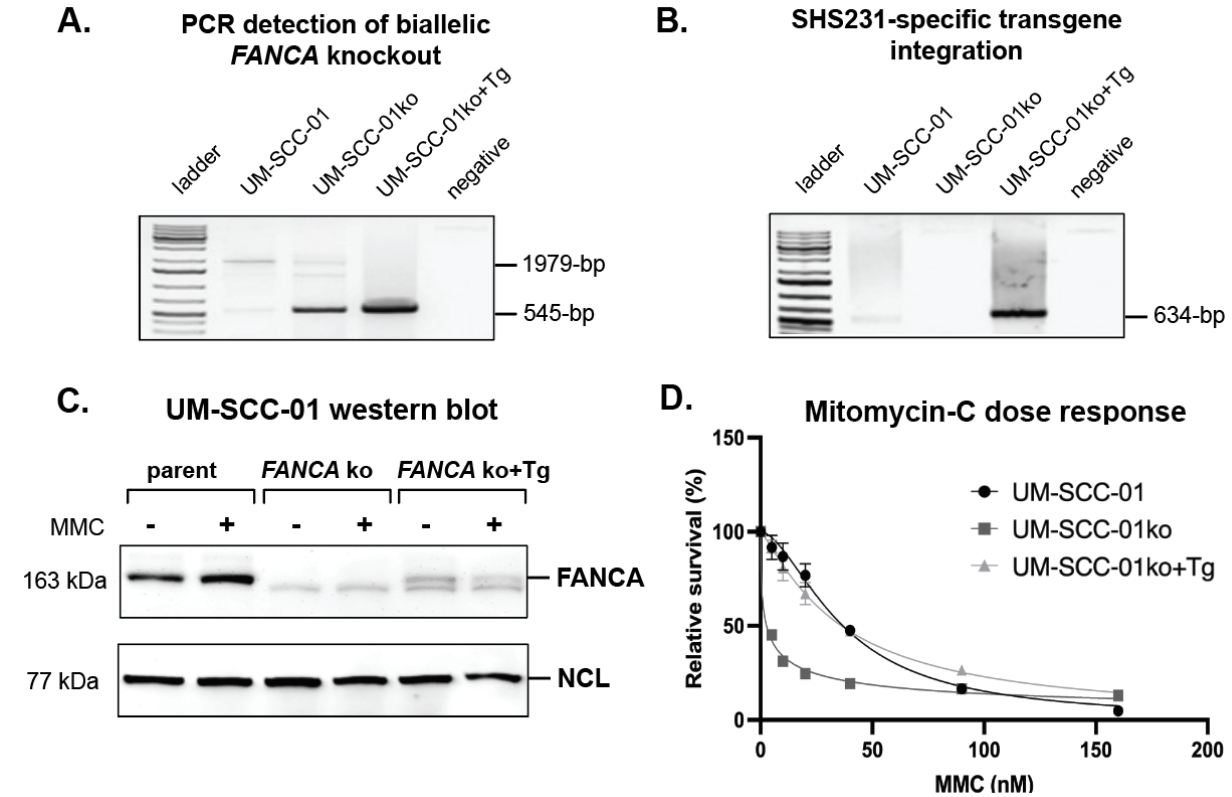


**Figure 1:** Workflow for generating, characterizing, and complementing *FANCA* isogenic cell line trios starting from sporadic HNSCC cell lines. (A) Generation of *FANCA* clonal knockouts. (B) Complementation of *FANCA* clonal knockouts.

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### Verifying *FANCA* KO and *FANCA* complemented clones

The following molecular, biochemical, and phenotypic assays were used to verify the generated *FANCA* knockout and *FANCA* complemented clone. A detailed description of the protocols to generate isogenic pairs and trios and results will be included in the manuscript.



**Figure 2:** Verification of *FANCA* KO and *FANCA* complemented clones. (A, B) Representative PCR results for detection of (A) wild type *FANCA* and dual gRNA/cas9 mediated *FANCA* ko alleles and (B) SH231 site specific insertion of *FANCA* complementation vector, pSH231-EF1-*FANCA*-Hygro. (C) Representative immunoblot result for detection of *FANCA* protein in 30  $\mu$ g of whole cell lysates. (D) Assessment of cell survival by alamarBlue assay 4 days after continuous exposure to mitomycin C (MMC).

### Protocols

#### 1. Molecular detection of *FANCA* wild type (WT) and knockout (KO) alleles:

Primers used:

- Forward primer sequence: 5' AATTGTTCTCCCGTCTGCTCTC 3'
- Reverse primer sequence: 5' GGGCCGTCTCCGTTAGTTTC 3'

#### PCR conditions:

- Denaturation: 94°C, 20 seconds
- Annealing: 56°C, 20 seconds
- Extension: 72°C, 2 minutes
- Number of cycles: 35

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Results: The two predicted PCR products using these primer sequences include a roughly 545 base pair deletion band (FANCA KO) and 1979 base pair intact band (FANCA WT). Both UM-SCC-01ko and UM-SCC-01ko+Tg cells will show the roughly 545 base pair deletion band, while the UM-SCC-01 cells will show the 1979 base pair wild-type intact band.

### 2. For detection of FANCA complementation vector in safe harbor site 231 (SH231):

Primers used:

- Forward primer sequence: 5' AGAACATGCAATGGCTAGC 3'
- Reverse primer sequence: 5' GCGGTGGTTGACCAGACAAA 3'

Results: The *FANCA* complemented, UM-SCC-01ko+Tg cells, display the predicted 634 base pair products, which represents of site-specific orientation.

### 3. Western blot of FANCA protein

Antibody used:

- FANCA primary antibody: Rabbit polyclonal (Bethyl laboratories #A301-980A)
- Secondary antibody: Goat anti-rabbit IgG, HRP conjugated (InVitrogen #G21234)

Results: UM-SCC-01 cells and FANCA complemented UM-SCC-01ko+Tg have a band indicating the presence of full-length wild type FANCA, whereas in UM-SCC-01ko cells have no detection of the full-length wild type FANCA protein due to the deletion in the *FANCA* gene.

### 4. Phenotypic analysis using a Mitomycin C sensitivity assay

Reagents:

- Mitomycin C (Millipore Sigma Cat. #M4287)
- alamarBlue Cell Viability Reagent (ThermoFischer Scientific DAL1025)

Procedure:

1. 2,500 cells are seeded per well in 48-well plates.
2. Cells are treated with varying concentrations of MMC ranging from 0nM to 80nM in fresh culture medium.
3. Incubate for 4 days.
4. Determine relative cell viability using the alamarBlue Cell Viability Reagent.

Results: UM-SCC-01ko demonstrate distinct hypersensitivity to MMC, while UM-SCC-01ko+Tg will show restored resistance comparable to wild type UM-SCC-01 cells.

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## References

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